

Technical Data Sheet



Specification

Solid differential and low water activity medium used for the determination of xerophilic fungi in low moisture food and in indoor air supplemented with trace metals.

Presentation

20 Prepared plates	Packaging Details	Shelf Life	Storage	
90 mm Plates	1 box with 2 packs of 10 plates/pack. Single	3 months	2-14°C	
with: 21 ± 2 ml	cellophane.			

Composition

Composition g:	
Peptone	5.000
Glucose	10.000
Potassium dihydrogen Phosphate	1.000
Magnesium sulfate	0.500
Dichloran	0.002
Chloramphenicol	0.100
Agar	15.000
Glycerol	175 ml
Trace Metal Solution	1.00 ml

Description /Technique

Description:

Among the culture media for xerophilic fungi, those that have played a more successful role are the ones which include any agent that restrains the continuous growth of zygomycete fungal colonies. Dichloran (dichlorebenzalkonium chloride) and Rose Bengal are two of those inhibitors.

DG18 Agar formulation used is that proposed by Hocking & Pitt in 1980, and it includes Dichloran which limits the size of fungal colonies more efficiently than Rose Bengal. Chloramphenicol inhibits bacterial growth and its thermostability allows it to be included in the medium before sterilization.

The inclusion of 18% (w/w) of Glycerine gives the medium a water activity (aw) of 0,955 without causing any of the problems that generally occur when this water activity is provided by sodium chloride or sugar.

The metal trace complement the inhibition and esporulation.

Technique:

Mass inoculation is recommended by spread plating using an inoculation loop, a swab or by spreading the sample with a Drigaslky loop. Never use an inoculum volume greater than 0,1 mL.

According to the standardized technique, plates must be incubated at 22-25°C, with partial readings after 3 and 5 days, and definitive readings after 7-8 days. Results are expressed in xerophiles-CFU/g or mL of food sample or CFU/m3 of air. Plates of DG18 Agar in bags will keep for up to one week at (5 ± 3)°C in the dark. Due to its extreme water activity (aw= 0.955), the plates must be rejected if any kind of dehydration is suspected.

Each laboratory must evaluate the results according to their specifications.

Presumptive isolation of any xerophile must be confirmed by further microbiological and biochemical tests.



Technical Data Sheet

Product: DG 18 METAL AGAR

Quality control

Physical/Chemical control

Color : Yellowish

pH: 5.6 ± 0.2 at 25°C

Microbiological control

Inoculate: Practical range 100 \pm 20 CFU; Min. 50 CFU (Productivity)/ 10^4 - 10^6 (Selectivity). Aerobic.Incubation at 22.5 \pm 2 flC until 5 days (moulds and yeast).

Microorganism	
---------------	--

Asperaillus brasiliensis ATCC[®] 16404, WDCM 00053 Escherichia coli ATCC[®] 25922, WDCM 00013 Bacillus subtilis ATCC[®] 6633, WDCM 00003 Candida albicans ATCC[®] 10231, WDCM 00054

Sterility Control

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH Check at 7 days after incubation in same conditions

Bibliography

· Atlas, R.M., et al. (1993) Handbook of Microbiological Media. CRC Press Inc. London.

• BEUCHAT, L.R. and C.A. HWANG (1995) Evaluation of modified dichloran 18% glycerol (DG18) agar for enumerating fungi in wheat flour. Int. J. Food Microbiol. 29:161-166.

· HOCKING, A.D. and J.I. PITT (1980) Dichloran-glycerol medium for enumeration of xerophilic funghi from low-moisture food. Appl. Environm. Microbiol. 39:488-492.

Growth

Good

Inhibited

Inhibited Good

. ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.

· ISO 16000-17 Standard. (2008) Indoor air.- Part 17: Detection and enumeration of moulds - Culture-based method.

· ISO 21527-2 :2008. Microbiology of food and animal feeding stuffs - Horizontal method for the enumeration of yeasts and moulds-Part 2: Colony count technique in products with water activity less than or equal to 0,95.

• PITT, J.I., and A.D. HOCKING (1985) Fungi and Food Spoilage. Academic Press. Sydney.

• PITT, J.I., A.D. HOCKING and D.R: GLENN (1983) An improved medium for the detection of Apergillus flavus and A. parasiticus. J. appl. Bacteriol. 54:109-114.

• SAMSON, R.A., E.S. HOEKSTRA, J.C. FRISVAD and O. FILTENBORG (2002) Introduction to the Food Borne Fungi. 6th ed. Centraalbureau voor Schimmelcultures. Utrech.

• TAPIA de DAZA, M.S. and L.R. BEUCHAT. (1992) Suitability of modified dichloran glycerol (DGH!8) agar for enumerating unstressed and stressed xerophilic molds. Food Microbiol. 9:319-333.