Condalab

Reference: 4531 **Technical Data Sheet**

Product: ROSE BENGAL CHLORAMPHENICOL THLTH AGAR

Specification

Solid and selective medium with neutralisers, for the isolation of yeasts and moulds, in surfaces.

Presentation

	Packaging Details	Shelf Life	Storage
30 Contact Plates/Ird. Contact Plates - Double Wrapping with: 15 ± 2 ml	1 box with 5 blisters (base of aluminium, PVDC and bag) with 6 contact plates/blister. Every pack exhibitis an irradiation indicator (8-14kGy).	5 months	2-25°C

Composition

Composition (g/l):

Mycological peptone	5.000
Dextrose	10.000
Potassium phosphate	1.000
Magnesium sulfate	0.500
Rose bengal	0.050
Chloramphenicol	0.100
Histidin	1.000
Lecithine	0.700
Polysorbate 80	5.000
Sodium Thiosulfate	0.500
Agar	15.000

Revision date: 07/02/19

Description /Technique

Description:

Rose Bengal Agar is a selective medium used to detect and enumerate moulds and yeasts in food samples. In addition the nutritional requirements for moulds and yeasts, this medium also contains Rose Bengal, which apart from turning the yeast a pink colour, facilitates counting, by reducing the luxuriant growth of moulds such as Rhizopus and Neurospora. This makes it is easier to detect other slower growing moulds.

The chloramphenicol included in the formulation inhibits bacterial growth, but does not interfere with the growth of fungi.

1.- The addition of he neutralizing agents TLTh (Tween 80 - Lecithin - Histidine - Sodium Thiosulphate) may inactivate a variety of disinfectants.

* The combination of lecithin, polysorbate 80 and histidine neutralizes aldehydes and phenolic compounds.
* The combination of lecithin and polysorbate 80 neutralizes the quaternary ammonium compounds.
* The polysorbate 80 neutralizes hexachlorophene and mercurial derivates.
* Sodium thiosulphate neutralizers halogen compounds.
* Lecithin neutralizes clorhexidine.
* Glycine: enhances the growth of microorganisms stressed

Technique

Contact plates are used in the microbiological control of disinfection and cleaning of surfaces. It acts simultaneously as a sampler and incubation culture medium without the need for any other intermediate steps.

The plates come in a form appropriate for this function and can be used with different culture media depending on the type of microbe that needs to be controlled. On average the plates provide a contact surface of approximately 25 cm2.

To use, remove the cover and gently press the culture medium on the surface to be controlled, ensuring contact between the two surfaces. The Contact plate is removed and covered with the lid to prevent air contamination. It is advisable that the lid is secured with adhesive tape and the bottom labelled with the sampling data (place, date and time). The inoculated plates are incubated at 25 ±1 ° C for 5 days and examined daily.

If the sample surfaces are rough, the Contact plates will not make good contact, even when the pressure is increased. In these cases it is advisable to delineate an sample surface area of 25 cm squared and rub this area vigorously with a wet sterile swab and then rub the swab over the Contact plate.

If verifying the effectiveness of a cleaning or disinfection process, Contact plates should be used within two hours after the end of the process, ensuring that the sample surface is dry. It is advisable to always include positive controls, sampling the area before disinfection or dirty areas beside the disinfected area.

The technician will determine the frequency of sampling and disinfection according to performance criteria.

Apply the agar directly onto surface to be monitorised ensuring that the pressure is distributed over the whole plate for 10 seconds.

Note: Contact plates are used for monitoring the microbiological contamination of surface and air inside cleanrooms, isolators, RABS, food industries and hospitals. The double/triple irradiated wrapping ensures that the package itself doesn't contaminate the environment as the first wrapper is removed just before entering the clean area.

Quality control

Physical/Chemical control

Color : Strongly pink pH: 7.2 ± 0.2 at 25°C

Microbiological control

Inoculate: Practical range 100 ± 20 CFU; Min. 50 CFU (Productivity)/ 10^4-10^6 (Selectivity).

Aaerobiosis. Incubation at 25°C±1, reading at 24-48-72 h to 5 days.

Microorganism	Growth
Candida albicans ATCC® 10231, WDCM 00054	Good (≥ 50 %)
Bacillus subtilis ATCC® 6633, WDCM 00003	Inhibited
Aspergillus brasiliensis ATCC® 16404, WDCM 00053	Good (≥ 50 %)
Escherichia coli ATCC® 8739, WDCM 00012	Inhibited

Sterility Control

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH
Check at 7 days after incubation in same conditions

Bibliography

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