

Reference: 4529 Technical Data Sheet

Product: ROSE BENGAL AGAR + CHLORAMPHENICOL

### **Specification**

Solid and selective medium for the isolation of yeasts and moulds from the environment and food products.

#### **Presentation**

30 Contact Plates/Ird. Contact Plates - Double Wrapping with: 15 ± 2 ml

### **Packaging Details**

1 box with 5 blisters (base of aluminium, PVDC and bag) with 6 contact plates/blister. Every pack exhibitis an irradiation indicator (8-14kGy).

Shelf Life

Storage

5 months 2-25°C

### Composition

Composition (g/l):	
Mycological peptone	5.000
Dextrose	10.000
Potassium phosphate	1.000
Magnesium sulfate	0.500
Rose bengal	0.050
Chloramphenicol	0.100
Agar	15.000

## **Description / Technique**

#### Description:

Rose Bengal Agar is a selective medium used to detect and enumerate moulds and yeasts in food samples. In addition the nutritional requirements for moulds and yeasts, this medium also contains Rose Bengal, which apart from turning the yeast a pink colour, facilitates counting, by reducing the luxuriant growth of moulds such as Rhizopus and Neurospora. This makes it is easier to detect other slower growing moulds.

The chloramphenical included in the formulation inhibits bacterial growth, but does not interfere with the growth of fungi.

### **Technique**

Contact plates are used in the microbiological control of disinfection and cleaning of surfaces. It acts simultaneously as a sampler and incubation culture medium without the need for any other intermediate steps.

The plates come in a form appropriate for this function and can be used with different culture media depending on the type of microbe that needs to be controlled. On average the plates provide a contact surface of approximately 25 cm2.

To use, remove the cover and gently press the culture medium on the surface to be controlled, ensuring contact between the two surfaces. The Contact plate is removed and covered with the lid to prevent air contamination. It is advisable that the lid is secured with adhesive tape and the bottom labelled with the sampling data (place, date and time). The inoculated plates are incubated at  $25 \pm 1$  ° C for 5 days and examined daily.

If the sample surfaces are rough, the Contact plates will not make good contact, even when the pressure is increased. In these cases it is advisable to delineate an sample surface area of 25 cm squared and rub this area vigorously with a wet sterile swab and then rub the swab over the Contact plate.

If verifying the effectiveness of a cleaning or disinfection process, Contact plates should be used within two hours after the end of the process, ensuring that the sample surface is dry. It is advisable to always include positive controls, sampling the area before disinfection or dirty areas beside the disinfected area.

The technician will determine the frequency of sampling and disinfection according to performance criteria.

Apply the agar directly onto surface to be monitorised ensuring that the pressure is distributed over the whole plate for 10 seconds.

Note: Contact plates are used for monitoring the microbiological contamination of surface and air inside cleanrooms, isolators, RABS, food industries and hospitals. The double/triple irradiated wrapping ensures that the package itself doesn't contaminate the environment as the first wrapper is removed just before entering the clean area.

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### **Quality control**

## **Physical/Chemical control**

Color : Strongly pink pH:  $7.2 \pm 0.2$  at  $25^{\circ}$ C

#### Microbiological control

Inoculate: Practical range 100 ± 20 CFU; Min. 50 CFU (Productivity)/ 10<sup>4</sup>-10<sup>6</sup> (Selectivity).

Aaerobiosis. Incubation at 25°C±1, reading at 24-48-72 h to 5 days.

Microorganism Growth

Candida albicans ATCC® 10231, WDCM 00054

Asperaillus brasiliensis ATCC® 16404, WDCM 00053

Escherichia coli ATCC® 8739, WDCM 00012

Bacillus subtilis ATCC® 6633, WDCM 00003

Good (≥ 50 %)

Inhibited

Inhibited

#### **Sterility Control**

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH Check at 7 days after incubation in same conditions

# **Bibliography**

- · ATLAS, R.M., L.C. PARKS (1993) Handbook of Microbiological Media. CRC Press, Inc. London.
- · CLESCERI, L.S., A.E. GREENBERG & A.D. EATON (1998) Standard Methods for the examination of water and wastewater. 20<sup>th</sup> ed. APHA. Washington DC.
- · DOWNES, F.P. & K. ITO (2001) Compendium of Methods for the Microbiological Examination of Foods. 4<sup>th</sup> ed. APHA. Washington DC.
- . ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- · MARSHALL, R.T. (1993) Standard methods for the examination of dairy products. 16th ed. APHA, Washington DC.

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